



Ahmad Hosseini-Safa, Ph.D.



Assistant Professor

Personal Data:

Date of Birth: 03/08/1986

Address (Work): Iran University of Medical Sciences, Shahid Hemmat Highway, Tehran, 1449614535, IRAN.

Telephone: +982186704601

Email: Hosseini.ahmad@IUMS.ac.ir – Ahmad.h.s65@gmail.com

Education:

- **2015- 2020:**

Doctor of Philosophy (Ph.D.) in Medical Parasitology, Tehran University of Medical Sciences, Tehran, Iran.

- **2012-2014:**

Master of Science degree (M.Sc.) in Medical Parasitology, Isfahan University of Medical Sciences, Isfahan, Iran.

- **2009-2011:**

Bachelor of Science (B.S.) in Medical Laboratory Science (MLS), Islamic Azad University of Medical Sciences, Shahroud, Iran.

- **2004-2007:**

Technician in Medical Laboratory Science (MLS), Islamic Azad University of Medical Sciences, Shahroud, Iran.

Language:

English:(MCHE: 64/100) (Internal exam).

Arabic: Native speaker.

Persian (Farsi): Native speaker.

H-index:

Google Scholar: 10

Scopus: 8

Researchgate: 10



Dissertations:

Ph.D.

Comparative study between molecular, parasitological, and immunology (avidity test) for diagnosing acute and chronic *Toxoplasma* infection in the animal model, pregnant women, and immunosuppressed patients.

M.Sc.

Genotyping of *Echinococcus granulosus* in slaughterhouse sheep infected with hydatid cyst in the Isfahan region according to the nucleotide sequence difference of the COX1 gene using the High-Resolution Melting (HRM) method.

Achievements:

1. Setting up the atomic force microscope (AFM) laboratory of Isfahan University of Medical Sciences.
2. Setting up a Real-Time PCR Laboratory, Department of Medical Parasitology and Mycology, School of Health, Tehran University of Medical Sciences.
3. The Head of Electron Microscopy Laboratory, including Atomic Force Microscopy (AFM) and the STM of the Comprehensive Research Laboratory, Isfahan University of Medical Sciences.
4. Lecturer of AFM microscope workshops at the Comprehensive Research Laboratory, Isfahan University of Medical Sciences.
5. Lecturer of Real-time PCR & HRM technique workshops of the Comprehensive Research Laboratory, Isfahan University of Medical Sciences.
6. 1st Ranked Graduated an associate degree in Medical Laboratory Sciences, 2007.
7. 1st Ranked Graduated Bachelor of Medical Laboratory Sciences (MLS), 2011.
8. 1st Ranked Graduated and member of the Association of Brilliant Talents and Elites in the Master's course of Medical Parasitology in Isfahan University of Medical Sciences, 2013.
9. 1st ranked in the interview for the Ph.D. exam with a score of 142 out of 150 total scores.
10. The fourth rank of the Ph.D. exam.
11. Member of Iranian Medical Parasitology Association.
12. Member of the Executive Committee of the First International Congress of New Medical Technologies, 2017.
13. Member of the Referrer Committee of the 10th International Laboratory and Clinical Congress, 2017.
14. The representative of the parasitology society of Iran in the Medical Council of I.R.IRAN, 2018.
15. The representative of the parasitology society of Iran on the student committee of the Society of Laboratory Sciences of Iran (for two years).
16. Member of the referrer Committee of the 11th International Laboratory and Clinical Congress, 2019.

Book:

- Treasure of Medical Parasitology, **Dr. Ahmad Hosseini-Safa**, and Dr. Mohammad Rostami-Nejad. Publisher: Teimourzadeh Novin. (ISBN: 9786006273907).

Research Interests:

1. Designing Real-time PCR kit
2. Molecular diagnosis methods
3. Molecular Parasitology
4. Protozoology
5. Parasite culture
6. Toxoplasmosis
7. Leishmaniasis

Technical skills:

1. PCR includes Conventional PCR RT-PCR and Real-Time PCR & HRM assay
2. DNA and RNA extraction
3. Primer and probe designing and analysis tools using Beacon Designer software, Primer Express Software, Primer Blast, Oligonucleotide Properties Calculator, and Oligo Analyzer software
4. Sequence-based database search (BLAST and FASTA)
5. Laboratory animal skills
6. Atomic force microscopy (AFM)
7. Tissue Samples section, preparation, and H&E Staining
8. ELISA
9. All standard Microsoft office software
10. Scientific and General Database Search

Journal Publication:

1. Pestechian, N., Tavakoli, S., Adibi, P., **Hosseini-Safa, A.**, Parsaei, R., & Yousefi, H. A. (2021). Prevalence of intestinal protozoan infection in patients with ulcerative colitis (UC) in Isfahan, Iran. *International Journal of Preventive Medicine*, 12.
2. Ouma, F. F., Nateghpour, M., Haghi, A. M., Mohebbali, M., Farivar, L., **Hosseini-Safa, A.**, & Mosawi, S. H. (2020). Application of High-Resolution Melting (HRM) Technique Towards the Detection of Asymptomatic Malaria in a Malaria Endemic Area of Southeastern Iran Under Elimination Program. *Journal of Arthropod-Borne Diseases*, 14(4), 353.
3. **Hosseini-Safa, A.**, Shojaee, S., Salami, S.A., Mohebbali, M., Hantoushzadeh, S., Mousavi, P., Manshadi, S.A.D. and Valian, H.K., 2020. Development of High-Resolution Melting Analysis as a Diagnostic Tool for Molecular Detection of *Toxoplasma* Infection in Pregnant Women and HIV Positive Cases. *Iranian Journal of Public Health*, 49(10), p.1983.

4. Allahmoradi, M., Haghi, A.M., Nateghpour, M., Mohebali, M., Raeisi, A., **Hosseini-Safa, A.**, Mohtasebi, S. and Afshar, M.J.A., 2020. High-Resolution Melting Analysis in Comparison with Microscopic Method: An Experimental Study to Diagnosis of Plasmodium Species Infections in Human. *Iranian Journal of Parasitology*, 15(3), p.403.
5. Sepahvand, **A.**, **Hosseini-Safa, A.**, Yousofi, H.A., Tajedini, M.H., Gharehbabah, R.P. and Pestehchian, N., 2020. Genotype Characteristics of *Giardia duodenalis* in Patients Using High-Resolution Melting Analysis Technique in Khorramabad, Iran. *Iranian Journal of Parasitology*, 15(2), p.204.
6. Moghadamizad, Z., **Hosseini-Safa, A.**, Mohebali, M., Heydarian, P., Aryaeipour, M. and Rokni, M.B., 2020. Specific detection of *Fasciola hepatica* and *F. gigantica* in infected domesticated animals using high-resolution melting analysis (HRM). *Iranian Journal of Public Health*, 49(3), pp.521-529.
7. Mosawi, S.H., Zarghona, Z., Dalimi, A., Jokelainen, P., **Hosseini-Safa, A.**, Mohammadi, M.R., Javanmardi, E. and Basirat, M.B., 2019. Particularly neglected in countries with other challenges: High *Toxoplasma gondii* seroprevalence in pregnant women in Kabul, Afghanistan, while a low proportion know about the parasite. *Plos one*, 14(10), p.e0223585.
8. **Hosseini-Safa, A.**, Rokni, M.B., Mosawi, S.H., Heydarian, P., Azizi, H., Davari, A. and Aryaeipour, M., 2019. High-resolution melting analysis as an appropriate method to differentiate between *Fasciola hepatica* and *F. gigantica*. *Iranian Journal of Public Health*, 48(3), p.501.
9. **Hosseini-Safa, A.**, Mohebali, M., Hajjaran, H., Akhoundi, B., Zarei, Z., Arzamani, K. and Davari, A., 2018. High resolution melting analysis as an accurate method for identifying *Leishmania infantum* in canine serum samples. *Journal of vector borne diseases*, 55(4), p.315.
10. **Hosseini-Safa, A.**, Mohag, M.A., Pestechian, N., Ganji, M., Mohammadi, R., Lamouki, R.M. and Rostami-Nejad, M., 2016. First report of Tasmanian sheep strain (G2) genotype isolated from Iranian goat using the high resolution melting (HRM) analysis. *Gastroenterology and Hepatology from Bed to Bench*, 9(Suppl1), p.S70.
11. Rostami-Nejad, R.N., Hejazi, S.H., Bahadoran, M., Pestehchian, N., Pourhoseingholi, M.A., **Hosseini-Safa, A.**, Sadeghi, A., Nobakht, H., Tajabadi Farahani, R. and Zali, M.R., 2016. Prevalence of *toxoplasma gondii* and the level of IL8 in patients with Celiac disease. *Koomesh*, pp.79-85.
12. Mousavi, S.M., **Hosseini-Safa, A.**, Mahmoudvand, H., Bahadorani, M.B., Mostafaei, S. and Darani, H.Y., 2016. No evidence of association between *Toxocara canis* infection and cancer risk. *Int J Med Res Health Sci*, 5(11), pp.204-208.
13. Sajjadi, S.E., Pestechian, N., Kazemi, M., Mohaghegh, M.A. and **Hosseini-Safa, A.**, 2016. Evaluation of the antimalarial effect of *Ferulago angulata* (Schlecht.) Boiss. extract and suberosin epoxide against *plasmodium berghei* in comparison with chloroquine using in-vivo test. *Iranian Journal of Pharmaceutical Research: IJPR*, 15(3), p.515.
14. **Hosseini-Safa, A.**, Mousavi, S.M., Badorani, M.B.B., Samani, M.G., Mostafaei, S. and Darani, H.Y., 2015. Seroepidemiology of toxocariasis in children (5–15 yr old) referred to the pediatric clinic of Imam Hossein Hospital, Isfahan, Iran. *Iranian journal of parasitology*, 10(4), p.632.

15. Ghomashlooyan, M., Vafaei, M.R., Kalani, H., Mirzaei, F., Azami, M., Jafari, R., Falahati, M., **Hosseini-Safa, A.** and Mohaghegh, M.A., 2015. Soil contamination with *Cryptosporidium* spp. in the west of Iran. *Parasitologists United Journal*, 8(2), p.123.
16. Mohammadi, R., **Hosseini-Safa, A.**, Ardakani, M.J.E. and Rostami-Nejad, M., 2015. The relation between intestinal parasites and some immune-mediated intestinal condition. *Gastroenterology and Hepatology from Bed to Bench*, pp.HepatoI-Bed.
17. **Hosseini-Safa, A.**, Harandi, M.F., Tajaddini, M., Rostami-Nejad, M., Mohtashami-Pour, M. and Pestehchian, N., 2015. Rapid identification of *Echinococcus granulosus* and *E. canadensis* using high-resolution melting (HRM) analysis by focusing on single nucleotide polymorphism. *Japanese Journal of Infectious Diseases*, pp.JJID-2015.
18. Mohammadi, R., Badiiee, P., Badali, H., Abastabar, M., **Hosseini-Safa, A.**, Hadipour, M., Yazdani, H. and Heshmat, F., 2015. Use of restriction fragment length polymorphism to identify *Candida* species, related to onychomycosis. *Advanced biomedical research*, 4.
19. Mohammadi, R., **Hosseini-Safa, A.**, Ardakani, M.J.E. and Rostami-Nejad, M., 2015. The relationship between intestinal parasites and some immune-mediated intestinal conditions. *Gastroenterology and hepatology from bed to bench*, 8(2), p.123.
20. Pestechian, N., **Hosseini-Safa, A.**, Tajedini, M., Rostami-Nejad, M., Mousavi, M., Yousofi, H. and Javanmard, S.H., 2014. Genetic diversity of *Echinococcus granulosus* in center of Iran. *The Korean Journal of Parasitology*, 52(4), p.413.
21. **Hosseini-Safa, A.**, Pestechian, N., Tajadini, M., Mohammadi, F., Cheraghipour, K. and Rostami Nejad, M., 2014. Evaluation the current status of hydatid cyst infection in slaughtered Livestock from Isfahan province. *Medical Science Journal of Islamic Azad Univesity-Tehran Medical Branch*, 23(4), pp.33-38.
22. Manesh, R.M., **Hosseini-Safa, A.**, Sharafi, S.M., Jafari, R., Bahadoran, M., Yousefi, M., Nasri, H. and Darani, H.Y., 2014. Parasites and chronic renal failure. *Journal of renal injury prevention*, 3(4), p.87.
23. Sharafi, S.M., Rostami-Nejad, M., Moazeni, M., Yousefi, M., Saneie, B., **Hosseini-Safa, A.** and Yousofi-Darani, H., 2014. *Echinococcus granulosus* genotypes in Iran. *Gastroenterology and hepatology from bed to bench*, 7(2), p.82.
24. Pestechian, N., Rasekh, H., Rostami-Nejad, M., Yousofi, H.A. and **Hosseini-Safa, A.**, 2014. Molecular identification of *Giardia lamblia*; is there any correlation between diarrhea and genotyping in Iranian population? *Gastroenterology and Hepatology from bed to bench*, 7(3), p.168.

Research Projects:

1. Seroepidemiology of toxocariasis in children (5–15 yr old) referred to the pediatric clinic of Imam Hossein Hospital, Isfahan, Iran (2011).
2. Genotyping of *Echinococcus granulosus* in slaughterhouse sheep infected with hydatid cyst in Isfahan province using High-Resolution Melting Analysis (HRM) (2012).
3. Molecular investigation of adult worms from camel isolates of *Echinococcus granulosus* in the Isfahan region using High-Resolution Melting Analysis (HRM) (2014).
4. Genotyping of *Giardia* spp. in patients referred to Khorram Abad laboratories using High-Resolution Melting Analysis (HRM) technique (2013).
5. Genotyping of *Echinococcus granulosus* in camel isolates of Isfahan region using High-Resolution Melting Analysis (HRM) (2014).
6. Investigating the effect of toxocariasis nematode egg antigens on eosinophilia in mice (2011).
7. Comparing the frequency distribution of anti-*Toxocara* antibody levels in the serum of cancer patients with healthy control groups (2011).
8. A comparative multigene study of *Leishmania infantum* genotypes using the High-Resolution Melting Analysis (HRM) method in domestic (dog) and wild (fox and jackal) reservoirs caught in Northeast Iran compared to the DAT test (2015).
9. Application of high-resolution melting (HRM) technique towards the detection of asymptomatic malaria in an endemic malaria area of Southeastern Iran under elimination program (2016).
10. Comparison of HRM and microscopy methods in laboratory diagnosis of *Plasmodium falciparum* and *Plasmodium vivax* and mixed infections (2016).
11. Seroprevalence of *Toxoplasma gondii* and associated risk factors among pregnant women in Kabul, the capital of Afghanistan (2018).
12. Genotyping of *Fasciola hepatica* and *Fasciola gigantica* adult worm in infected livestock isolates from Lorestan province using high-resolution melting (HRM) (2018).
13. Simultaneous identification of four bacteria causing meningitis (*Neisseria meningitidis*, *Streptococcus pneumoniae*, *Streptococcus agalactiae*, and *Hemophilus influenzae*) in suspected meningitis patients using the High-Resolution Melting Analysis (HRMA) method (2017).
14. Evaluation of the Semi-Nested Real-Time PCR MGB Taqman Probe method in diagnosing acute and chronic *Toxoplasma* infection in an animal model; Pregnant women and immune-compromised patients (2018).
15. Assessment of IgG avidity ELISA test to discriminate acute and chronic stages of visceral leishmaniasis in domestic dogs as potential reservoir hosts of Mediterranean type of kala-azar (2019).

Certificates:

1. International certificate of laboratory management and quality control (ISO/IEC17025) QAL UK license.
2. International Certificate of BioTek company in Cytation 3, Multimode Cell Imaging Reader.
3. Certificate of Real-Time PCR Workshop, Applied Physiology Research Center, Isfahan University of Medical Sciences.
4. Certificate of Primer & Probe Design workshop, Applied Physiology Research Center, Isfahan University of Medical Sciences.
5. Certificate of Bioinformatics and Method in Biomedical Research workshop, Applied Physiology Research Center, Isfahan University of Medical Sciences.
6. Certificate of Scientific Written Workshop, School of Public Health, Tehran University of Medical Sciences.
7. Certificate of EndNote software workshop, School of Public Health, Tehran University of Medical Sciences.
8. Workshop on teaching strategies and methods, Faculty of Health, Tehran University of Medical Sciences.
9. Oral presentation at the 3rd International Congress on Parasitic Diseases, 2017.
10. Oral presentation at the 1st International Congress of Modern Medical Technologies, 2017.
11. Two oral presentations at the 4th International Congress on Parasitic Diseases, 2019.

Congress:

1. 5th National Conference of Iranian Medical Sciences Jan 2007.
2. 1st International & 8th National Congress of Parasitology & Parasitic Diseases in Iran Oct 16-18, 2012, Kerman.
3. 6th International Congress of Laboratory & Clinic, 2013.
4. 13th International Congress of Parasitology. Mexico, 2014.
5. 2nd International & 10th National Congress on Parasitology & Parasitic Diseases of Iran, Gilan, 2015.
6. 3rd International & 10th National Congress on Parasitology & Parasitic Diseases of Iran, Shiraz, 2017.
7. 5th National & 1st International Congress on Novel & Innovative Laboratory Technologies Razi International Conference Center, 2017.
8. 10th International Congress of Laboratory & Clinic, 2017.
9. Chromosome-Centric Human Proteome Project Symposium (C-HPP).
10. 11th International Congress of Laboratory & Clinic, 2019.
11. 10th International Congress of Laboratory & Clinic, 3rd National Congress of Basic Medical Sciences and Knowledge-Based Production, 2019.
12. 4th International & 10th National Congress on Parasitology & Parasitic Diseases of Iran, Urmia, 2019.

